

Insulin-like growth factor 1 (IGF-1) pathway targeting in the treatment of cancer

L. Decoster, E. Menu, K. Vanderkerken, J. De Grève

The insulin-like growth factor 1 (IGF-1) pathway has been under therapeutic investigation for many years, both in cancer and in other diseases. This signaling pathway plays an important role in the normal growth and development of vertebrate organisms, but has limited function in adults. At the cellular level it is fundamental in a variety of processes including cell proliferation and survival.^{1,2} As such, the pathway plays a major role in tumour cell biology, including proliferation and survival, making it an apparently attractive target for therapeutic intervention. IGF1 is ubiquitously expressed in all organs with extreme high levels (compared to all other organs) found in the normal prostate. This review aims to give a short overview of reported clinical trials with drugs targeting the source of the pathway and in particular efforts targeting the Insulin-like Growth Factor Receptor-1 (IGF-1R).

(*Belg J Med Oncol* 2011;5:108-15)

Anatomy of the IGF-1 pathway

The initiating elements of the IGF-1 pathway are 3 ligands, IGF-1, IGF-2 and insulin; 3 cell membrane receptors, IGF-1R, Insulin receptor (IR) and IGF-2 receptor; and 6 high-affinity IGF binding proteins (IGFBP). IGF-I and IGF-II are found in the circulation, complexed to IGFBP's, which serve to regulate bioavailability of these ligands in the tissue.

The IGF-1R is a receptor tyrosine kinase functioning as a heterotetramer of 2 extracellular ligand binding α subunits and 2 β subunits comprising the transmembrane and tyrosine kinase domains.² It shares 60% homology at the amino acid sequence level with the insulin receptor (IR).³ The IGF-1R has high affinity for both IGF-1 and IGF-2. Binding of the ligands to the IGF-1R leads to the de-inhibition of the intrinsic

tyrosine kinase activity, which results in autophosphorylation of tyrosine residue. This leads to binding of substrates and initiation of phosphorylation cascades that serve to transmit the IGF-1R signal. As a result 2 distinct pathways are activated: the PI3K-Akt-mTOR pathway which predominantly stimulates cell survival and the RAS-RAF-MAPK, which predominantly mediates cell proliferation (*Figure 1*).^{2,4,5}

Implication of the IGF-1 pathway in malignancy

The IGF-1R and its 2 ligands play a critical role during embryogenesis and development in humans. In general, serum IGF-1 levels decline progressively after puberty. The implication of the pathway in can-

Authors: Ms. L. Decoster MD Department of Medical Oncology, University Hospital Brussels, Brussels; Ms. E. Menu MD PhD; Ms. K. Vanderkerken MD PhD Department of Hematology and Immunology, Vrije Universiteit Brussel, Brussels; Mr. J. De Greve, MD PhD, Department of Medical Oncology, University Hospital Brussels, Brussels.

Please send all correspondence to: Ms. L. Decoster, Department of Medical Oncology, University Hospital Brussels, Laarbeeklaan 101, Brussels, Belgium, Email: lore.decoester@uzbrussel.be.

Conflict of interest: the authors have nothing to disclose and indicate no potential conflicts of interest.

Key words: IGF-1 receptor

cer can be considered in itself or in its relationship with other oncogenic pathways.

The IGF-1 pathway

Increased expression of IGF-1, IGF-2, IGF-1R, or combinations have been documented in various malignancies, including breast cancer, ovarian cancer, malignancies of the gastrointestinal tract such as colorectal cancer, hepatocellular carcinomas, pancreatic carcinomas and haematological malignancies.⁶⁻⁹ In addition, epidemiological studies have shown that high levels of serum IGF-1 and/or lower levels of IGFBP's are associated with an increased risk for several cancers, including premenopausal breast cancer, prostate cancer, lung cancer and colorectal cancer.¹⁰

In vivo, overexpression of the IGF-1R accelerated the progression of pancreatic cancer in a mouse model, mainly by promoting the capability for invasion and metastasis.¹¹

The functional and clinical prognostic significance of the IGF-1R expression and of its ligand levels has already been demonstrated for some malignancies (such as multiple myeloma) but still needs to be determined for most.

So far, the relative overexpression of the IGF-1R in a tumour compared to normal tissue is shown in a number of studies, e.g. in colon cancer.¹² The constitutive nature and, as a consequence, the importance of such overexpression for the pathogenesis and progression of most human cancers remains to be determined. Only in a few examples evidence of constitutive activation of the pathway is available. In some sarcomas, ligand overexpression seems to be constitutively driven by genomic alterations in the tumour. In Ewing sarcoma the characteristic t (11; 22) chromosomal translocation resulting in the EWS-FLI1 gene fusion has been shown to cause up-regulation of IGF-1 and down-regulation of IGFBP-3 causing autocrine stimulation of the IGF-1R.¹³ Synovial sarcomas exhibit characteristic t (X; 18) translocations resulting in enhanced transcription of the IGF-2 gene and hyperactivation of IGF-1R signaling.¹⁴ Amplification of the IGF-1R gene has been found infrequently in breast cancer (<2%) and melanoma.^{15,16} In small cell lung cancer, some low level amplification or polysomy was found, but did not affect prognosis.¹⁷ In non-small cell lung cancer,

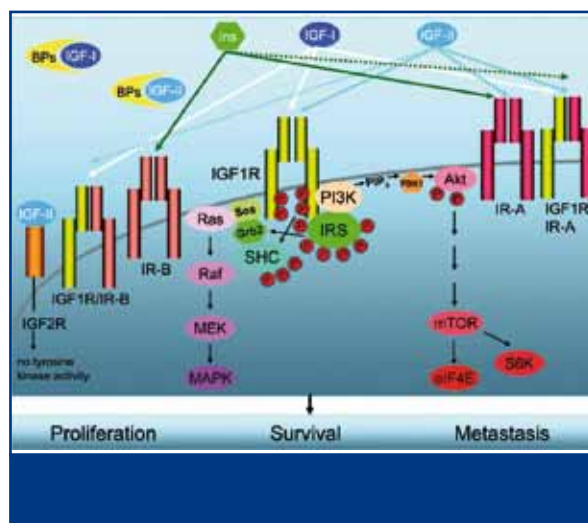


Figure 1. The IGF pathway (adapted from Sachdev D and Yee D. Disrupting insulin-like growth factor signaling as a potential cancer therapy.⁵)

gene amplification was found in 2.8%.¹⁸ In addition, IGF-1R protein and gene expression were not associated with survival, whereas high IGF-1R gene copy number had a *positive* prognostic value.¹⁴ Mutations of the receptor have been described only in a few human cancer cases (brain, lung, renal) in genome wide sequencing efforts.¹⁹ In Multiple Myeloma (MM), IGF-1R expression seems to be linked to t (4; 14) translocations and to the expression of certain cell markers.⁹ CD45 phosphatase inhibits IGF-1 signaling while CD56 positive myeloma cells reacted to IGF-1, whereas CD56 negative cells did not.^{20,21} The functional consequences of direct or indirect constitutive activation of the pathway and value for therapeutic targeting remain to be defined further.

In relationship to other oncogenic pathways

The IGF-1 pathway has also been shown to cross-talk with a number of other signaling pathways and is involved in feedback loops that are activated when other, activated, pathways are therapeutically inhibited. For example, functional up-regulation of IGF-1R signaling has recently been implicated in the development of resistance to anti-cancer therapy such as human epidermal growth factor receptor 2 (HER2) targeting and EGFR targeting.^{22,23} This mechanism could be a rationale for combined targeting of the

EGFR or HER2 on the one hand and IGF-1R on the other hand. In MM, a cross-talk exists between the IL-6 and the IGF-1 pathway, with IGF-1 enhancing IL-6 effects.²⁴ In addition, IGF-1R signaling has also been implicated in resistance to endocrine therapies in breast cancer, radiotherapy and a variety of chemotherapy agents, providing additional rationale for the investigation of combinatorial strategies.

Therapeutic targeting the IGF-1 pathway in cancer

The IGF pathway could thus be considered as a therapeutic target on its own or as an additional target to help other targeted therapies.

The IGF-1R is thought to be of potential therapeutic value because of its importance to cell growth, cell metabolism and cell survival and some hope that combining IGF-1R inhibition might enhance the activity of other targeted therapies, chemotherapy or radiotherapy. In vitro and in vivo inhibition of IGF-1R signaling in unselected cancer cells resulted in apoptosis in malignant cells and inhibition of tumour formation.²⁵

However, based on expanding clinical evidence with several other therapeutic molecular targets, it is to be expected that the IGF-1R as a primary therapeutic target on its own would be specifically worthwhile to be investigated only in instances in which the pathway is constitutively activated through mechanisms elaborated above or yet to be discovered.

Strategies to inhibit the IGF-1R pathway include on the one hand the reduction of ligand levels or activity and on the other hand inhibition of receptor function using anti-receptor antibodies or small molecule tyrosine kinase inhibitors.

As primary target

An attempt to reduce IGF-1 levels using a somatostatin analogue did not result in the intended declines in IGF-1 or insulin levels in early stage breast cancer patients.²⁶ Anti-IGF-1 antibodies have shown interesting preclinical activity and are being investigated further.²⁷

Antibodies targeting the IGF-1R that are very selective for IGF-1R are available. Binding of IGF-1R antibodies to the receptor prevents ligand-induced activation and induces receptor internalisation and

degradation by endocytosis. Most of the known antibodies are fully human or humanised IgG1 antibodies, except figitumumab which is an IgG2 antibody.²⁸

In MM, several IGF-1 antibodies have been pre-clinically tested with encouraging results: Imclone's IMC-A12, Pfizer's CP-751,871 and Merck's Dalotuzumab all show anti-tumour effects in murine models.²⁹⁻³¹ Tyrosine kinase inhibitors (TKI's) inhibit receptor activation by directly binding to and blocking the catalytic kinase domain. However, TKI's are less selective for the IGF-1R because of the homologies with the IR at the amino acid level, which may result in greater potency but also significantly increased toxicity due to co-targeting of the IR.³² Despite their limited selectivity, several TKI's have been investigated preclinically in several cancer types including MM: NVP-ADW742 and NVP-AEW541 from Novartis, PPP or BVP-51004 from Biovitrium and GSK1838705A and GSK1904529A from GlaxoSmithKline.^{7,33-36} All show promising anti-tumoural effects.

Targeting IGF-1 pathway in combination therapies

Several preclinical studies have been performed, for example in MM to analyse the effects of combining anti-IGF-1R therapy with existing conventional drugs. Rapamycin (an mTOR inhibitor) has been shown to work synergistically with IMC-A12 to inhibit tumour growth in vitro and IMC-A12 has also been shown to synergistically reduce tumour growth in a xenograft model when combined with bortezomib and/or melphalan.²⁹ NVP-ADW742 combined with melphalan synergistically reduces in vivo tumour growth and NVP-AEW541 enhances the effects of lenalidomide and bortezomib in vitro.^{7,32} Recently NVP-AEW541 has also been combined with a novel mTOR inhibitor, namely Rad001 to reduce myeloma growth in vitro.³⁸

Clinical trials

Phase I trials

As primary target

Phase I trials with different drugs targeting the IGF-1R have revealed a favourable profile with most adverse events described as mild. The most common

Table 1. Summary of clinical trial results with anti-IGF-1R antibodies

Agent	Dosing schedule	Trial Results
CP-751,871 (figitumumab) fully human IgG2	q3-4 weeks intravenous	Phase I: EWS 1CR STS 12/20 SD adrenal carcinoma 9/13 SD MM +dexamethasone 9/27PR CRPC + docetaxel: 8/20 PR Phase II: NSCLC + carboplatin/paclitaxel: OR 54% Phase III: NSCLC carboplatin/paclitaxel +/- figitumumab: closed prematurely, no benefit and increased toxicity
A12 (cixutumumab) fully human IgG1	q2 weeks intravenous	Phase I: SD>9 months in 2 pts (1 male breast cancer; 1 HCC) Phase II: CRPC: SD>6 mo in 29% CRC +/- cetuximab: insufficient activity
AMG-479 (ganitumab) fully human IgG1	q2 weeks intravenous	Phase I: EWS: 1CR carcinoid tumor: 1PR CRC refractory to cetuximab: +panitumumab 1PR Phase II EWS or desmoplastic small round cell tumors: OR 6%; OR+SD>24weeks 20% pancreas carcinoma + Gemcitabine: trend towards longer PFS, longer 6 months OS; more SD
R1507 (pobatumumab) fully human IgG1	q1-3 weeks intravenous	Phase I: EWS: 2PR Phase II: sarcoma: activity observed in EWS, rhabdomyosarcoma and osteosarcoma EWS: OR 14.4%
MK0646 (dalotuzumab) humanized IgG1	q1-2week intravenous	Phase I: solid tumors: 2 pts with SD>1 year; 3 pts with metabolic response Phase II: NET: insufficient activity

EWS: Ewing sarcoma; CR: complete remission, STS: soft tissue sarcoma; SD: stable disease; PR: partial remission; MM: multiple myeloma; CRPC: castration resistant prostate cancer; pts: patients; HCC: hepatocellular carcinoma; CRC: colorectal cancer; PFS: progression free survival; OS: overall survival; NET: neuroendocrinal tumour

adverse events seem to be due to endocrine changes, including hyperglycaemia. Co-administration of anti-diabetic agents may be necessary to control blood glucose. Up to 20% of severe hyperglycaemia is seen in studies in which anti-IGF-1R antibodies are combined with chemotherapy regimen that require corticosteroids as premedication.³⁹ Haematological toxicity including grade 3 thrombocytopenia has been observed mainly with IgG1 antibodies.^{40,41} Hypersensitivity reactions were rare.^{32,42}

In different phase I trials objective responses have been reported in patients with Ewing sarcoma: 1 complete remission (CR) with AMG-479⁴⁰, 1 CR

with CP-751,871 (figitumumab) and 2 objective responses (OR) and 2 stable disease (SD) with R1507 (robatumumab).^{40,43,44} Figitumumab also showed signals of activity in sarcoma (12 patients with disease stabilisation) and in prostate cancer (PSA responses in combination with docetaxel) and in MM 9/27 patients with relapsed disease responded while no dose-limiting toxicities were identified.^{41,45,46} Dalotuzumab is currently also in phase I clinical trial for various solid tumours and MM.³¹ Two previous phase I trials in advanced solid tumours demonstrated metabolic response in 3 patients and SD lasting for over a year in 2 patients.^{47,48}

A phase I trial with OSI-906, a small-molecule dual inhibitor of IGF-1R/IR, demonstrated stable disease lasting for over 12 weeks in 7/20 patients, including 1 thymus carcinoma, 1 adrenal carcinoma and 1 colorectal carcinoma.⁴⁹

Targeting IGF pathway in combination therapies

For example, in MM, a phase I trial has been completed combining IMC-A12 with temsirolimus (a rapamycin analogue) in patients with advanced disease and a phase I trial is ongoing combining the TKI AVE-1642 (Sanofi-Aventis) with bortezomib in MM patients with relapsed disease, results are not available yet (source: NIH).

Phase II trials

As primary therapeutic target

A phase II trial of R1507 in patients with recurrent or refractory sarcoma reported clinical significant activity in Ewing sarcoma, rhabdomyosarcoma and osteosarcoma.⁵⁰ The same monoclonal antibody was investigated in patients with recurrent or refractory Ewing sarcoma family of tumours and demonstrated an OR in 14.4% with 12/125 patients responding at least 6 weeks with a median duration of 25 weeks.⁵¹ Another phase II trial with AMG-479 in Ewing sarcoma and desmoplastic small round cell tumours demonstrated an OR in 6% (2 patients with PR) but a clinical benefit (PR+ SD lasting more than 24 weeks) of 20%.⁵² In colorectal cancer refractory to cetuximab or panitumumab, cixutumumab with or without cetuximab did not show any activity.⁵³ A phase II trial of cixutumumab monotherapy in castration refractory prostate cancer suggested modest antitumour activity with disease stabilisation for over 6 months in 29% of patients.⁵⁴ In prostate cancer the administration of preoperative figitumumab demonstrated biological activity with PSA decline.⁵⁵ The activity in prostate cancer is intriguing, as a specific mechanism of genomic activation of the pathway in prostate cancer has not been documented yet. Perhaps this could be related to the high importance of IGF-1 in the normal prostate or a yet to be defined constitutive driver of the pathway which might very well include the driving of the IGF-1 pathway due to constitutive activation (by gene rearrangement) of Ets and Erg in an important fraction of prostate cancers.⁵⁶

Targeting the IGF pathway to help other targeted therapies

In the virtual absence of the current identification of sensitising mutations in the IGF-1 pathway in human cancer, a major application for IGF-1 pathway targeting might reside in augmenting the therapeutic effect of targeting other activated oncogenic pathways.

A phase II trial with figitumumab in combination with paclitaxel and carboplatinum as first line treatment in patients with locally advanced or metastatic non-small cell lung cancer reported a promising OR of 54% versus an OR of 42% in the chemotherapy alone arm.⁵⁷ The response rates were the highest in patients with squamous cell carcinoma. Based on these results, a randomised phase III trial was initiated (see below).

Another phase II trial adding AMG-479 to gemcitabine in metastatic pancreatic cancer, demonstrated a trend toward longer progression free survival, 6 months overall survival and higher rates of stable disease.⁵⁸

Phase III trials

In NSCLC a randomised phase III trial compared the combination of figitumumab to carboplatinum and paclitaxel versus carboplatinum and paclitaxel alone. This trial was suspended after the accrual of 681 of the planned 820 patients because an interim analysis showed no improvement of the efficacy over chemotherapy alone and an increase in toxicity.⁵⁹ A similar negative experience has been made in breast cancer reported recently at the San Antonio Breast Cancer Conference.⁶⁰

Both clinical development strategies have suffered from the failure to preselect patients at least for the presence of expression of the targeted protein. It is most unfortunate that such costly efforts (that also demand active participation of patients) due to these design limitations risk throwing out erroneously some possibly valuable treatments for a subset of patients.

Conclusions

Preclinical and early clinical results have suggested potential utility of IGF-1R targeting agents in the management of cancers. Promising preclinical work and even phase II data have however not led to further validation of the therapeutic strategy as yet, except for 2 cancers in which the IGF-1 pathway might be indirectly constitutively activated: Ewing sarcoma and prostate cancer. Patient selection may

be the key to success in developing this therapeutic strategy further. Indeed, in other cancers, the most striking therapeutic effects have been obtained when the target gene itself is genomically activated, by mutation (e.g. EGFR in lung cancer, c-KIT in GIST) or translocation (bcr-abl in CML, Alk fusion gene in lung cancer). Significant but slightly less impact is observed in cases in which the target gene is amplified (HER2 in breast cancer, gastric cancer). In contrast, when the target gene is in its wild type status, minimal or no therapeutic benefit can be observed (e.g. wild type EGFR in lung cancer). When the therapeutic target is indirectly activated by an upstream driver (e.g. VEGF in renal cell cancer), a therapeutic benefit can be obtained, but in the cases at hand, of less magnitude than when the target gene itself is activated.

The minority of breast and lung cancers that have some IGF-1R gene amplification could therefore be explored for the therapeutic efficacy of this anti-cancer strategy. Similarly, as a secondary target, this strategy could be investigated further in combination with other targeted agents in populations with a genomically activated primary oncogenic pathway (e.g. mutant EGFR in lung cancer or HER2 amplified breast cancer), but at least with proper selection of patients with regard to expression of the IGF-1R.

References

1. Samani AA, Yakar S, LeRoith D, Brodt P. The role of the IGF system in cancer growth and metastasis: overview and recent insights. *Endocr Rev* 2007;28:20-47.
2. Hartog H, Wesseling J, Boezen HM, van der Graaf WTA. The insulin-like growth factor 1 receptor in cancer: old focus, new future. *Eur J Cancer* 2007;43:1895-1904.
3. Baserga R, Hongo A, Rubini M, Prisco M, Valentis B. The IGF-I receptor in cell growth, transformation and apoptosis. *Biochim Biophys Acta* 1997;1332:F105-26.
4. Chitnis MM, Yuen JSP, Protheroe AS, Pollak M, Macaulay VM. The type 1 insulin-like growth factor receptor pathway. *Clin Cancer Res* 2008;14:6364-70.
5. Sachdev D, Yee D. Disrupting insulin-like growth factor signaling as a potential cancer therapy. *Mol Cancer Ther* 2007;6:1-12.
6. Chng, W J, Gualberto A, Fonseca R. IGF-1R is overexpressed in poor-prognostic subtypes of multiple myeloma. *Leukemia* 2006;20:174-6.
7. Mitsiades CS, Mitsiades NS, McMullan CJ, Poulaki V, Shringarpure R, Akiyama M, et al. Inhibition of the insulin-like growth factor receptor-1 tyrosine kinase activity as a therapeutic strategy for multiple myeloma, other hematologic malignancies, and solid tumours. *Cancer Cell* 2004;5:221-30.
8. Standal T, Borset M, Lenhoff S, Wisloff F, Stordal B, Sundan A, et al. Serum insulinlike growth factor is not elevated in patients with multiple myeloma but is still a prognostic factor. *Blood* 2002;100:3925-9.
9. Sprynski AC, Hose D, Caillot L, Réme T, Shaughnessy JD Jr, Barlogie B, et al. The role of IGF-1 as a major growth factor for myeloma cell lines and the prognostic relevance of the expression of its receptor. *Blood* 2009;113:4614-26.
10. Furstenberger G, Senn HJ. Insulin-like growth factors and cancer. *Lancet Oncol* 2002;3:298-302.
11. Lopez T, Hanahan D. Elevated levels of IGF-1 receptor convey invasive and metastatic capability in a mouse model of pancreatic islet tumorigenesis. *Cancer Cell* 2002;1:339-53.
12. Weber MM, Fottner C, Liu SB, Jung MC, Engelhardt D, Baretton GB. Overexpression of the insulin-like growth factor I receptor in human colon carcinomas. *Cancer* 2002;95:2086-95.
13. Balamuth NJ, Womer RB. Ewing's sarcoma. *Lancet Oncol* 2010;11:184-92.
14. Friedrichs N, Kuchler J, Endl E, Koch A, Czerwitzki J, Wurst P, et al. Insulin-like growth factor-I receptor acts as a growth regulator in synovial sarcoma. *J Pathol* 2008;216:428-39.
15. Berns EM, Klijn JG, Van Staveren IL, Portegen H, Foekens JA. Sporadic amplification of the insulin-like growth factor 1 receptor gene in human breast tumours. *Cancer Res* 1992;52:1036-9.
16. Zhang N, Trent JM, Meltzer PS. Rapid isolation and characterization of amplified DNA by chromosome microdissection: identification of IGF-1R amplification in malignant melanoma. *Oncogene* 1993;8:2827-31.
17. Badzio A, Wynes MW, Dziadziuszko R, Merrick D, Pardo M, Singh S, et al. Protein expression (PE) and increased IGF-1R gene copy number (GCN) in small cell lung cancer (SCLC). *J Clin Oncol* 2010;28 Suppl 15:10584.
18. Dziadziuszko R, Merrick DT, Witta SE, Mendoza AD, Szostakiewicz B, Szymanowska A, et al. Insulin-like growth factor receptor 1 (IGF1R) gene copy number is associated with survival in operable non-small-cell lung cancer: a comparison between IGF-1R fluorescent in situ hybridization, protein expression and mRNA expression. *J Clin Oncol* 2010;28:2174-80.
19. <http://www.sanger.ac.uk/perl/genetics/CGP/cosmic?action=gene&ln=IGF-1R>
20. Descamps G, Willeme-Toumi S, Trichet V, Venot C, Debussche L, Hercend T, et al. CD45neg but not CD45pos human myeloma cells are sensitive to the inhibition of IGF-1 signaling by a murine anti-IGF-1R monoclonal antibody,

mAVE1642. *J Immunol* 2006;177: 4218-23.

21. Sahara N, Takeshita A, Ono T, Sugimoto Y, Kobayashi M, Shigeno K, et al. Role for interleukin-6 and insulin-like growth factor-I via PI3-K/Akt pathway in the proliferation of CD56- and CD56+ multiple myeloma cells. *Exp Hematol* 2006;34:736-44.

22. Nahta R, Yu D, Hung MC, Hortobagyi GN, Esteva FJ. Mechanisms of disease: understanding resistance to HER2-targeted therapy in human breast cancer. *Nat Clin Pract Oncol* 2006;3:269-80.

23. Chakravanti A, Loeffler JS, Dyson NJ. Insulin-like growth factor receptor I mediates resistance to anti-epidermal growth factor receptor therapy in primary human glioblastoma cells through continued activation of phosphoinositide 3-kinase signaling. *Cancer Res* 2002;62:200-7.

24. Jelinek DF, Witzig TE, Arendt BK. A role for insulin-like growth factor in the regulation of IL-6-responsive human myeloma cell line growth. *J Immunol* 1997;159:487-96.

25. Resnicoff M, Coppola D, Sell C, Rubin R, Ferrone S, Baserga R. Growth inhibition of human melanoma cells in nude mice by antisense strategies to type 1 insulin-like growth factor receptor. *Cancer Res* 1994;54:4848-50.

26. Pollak MN, Chapman JW, Pritchard KI, Krook JE, Dhaliwal HS, Vandenberg TA, et al. Tamoxifen versus tamoxifen plus octreotide LAR as adjuvant therapy for early-stage breast cancer in postmenopausal women: Update of NCIC CTG MA14 trial. *J Clin Oncol* 2010;28 Suppl 15:542.

27. Goya M, Miyamoto S, Nagai K, Ohki Y, Nakamura K, Shitara K, et al. Growth inhibition of human prostate cancer cells in human adult bone implanted into nonobese diabetic/Severe combine immunodeficient mice by a ligand-specific antibody to human insulin-like growth factors. *Cancer Res* 2004;64:6252-8.

28. Gualberto A, Karp DD. Development of the monoclonal antibody figitumumab, targeting the insulin-like growth factor-1 receptor, for the treatment of patients with non-small-cell lung cancer. *Clin Lung Cancer* 2009;10:273-80.

29. Wu KD, Zhou L, Burtrum D, Ludwig DL, Moore MA. Antibody targeting of the insulin-like growth factor I receptor enhances the anti-tumour response of multiple myeloma to chemotherapy through inhibition of tumour proliferation and angiogenesis. *Cancer Immunol Immunother* 2007;56:343-57.

30. Cohen BD, Baker DA, Soderstrom C, Tkalecivic G, Rossi AM, Miller PE, et al. Combination therapy enhances the inhibition of tumour growth with the fully human anti-type 1 insulin-like growth factor receptor monoclonal antibody CP-751,871. *Clin Cancer Res* 2005;11:2063-73.

31. Scartozzi M, Bianconi M, Maccaroni E, Giampieri R, Berardi R, Cascinu S. Dalotuzumab, a recombinant humanized mAb targeted against IGFR1 for the treatment of cancer. *Curr*

Opin Mol Ther 2010;12:361-71.

32. Zha J, Lackner MR. Targeting the insulin-like growth factor receptor-1R pathway for cancer therapy. *Clin Cancer Res* 2010;16:2512-7.

33. Maiso P, Ocio EM, Garayoa M, Montero JC, Hofmann F, Garcia-Echeverria C, et al. The insulin-like growth factor-I receptor inhibitor NVP-AEW541 provokes cell cycle arrest and apoptosis in multiple myeloma cells. *Br J Haematol* 2008;141:470-82.

34. Menu E, Jernberg-Wiklund H, Stromberg T, De Raeve H, Girnita L, Larsson O, et al. Targeting the IGF-1R using picropodophyllin in the therapeutic 5T2MM mouse model of multiple myeloma: beneficial effects on tumour growth, angiogenesis, bone disease and survival. *Int J Cancer* 2007;121:1857-61.

35. Sabbatini P, Korenchuk S, Rowand JL, Groy A, Liu Q, Leperi D, et al. GSK1838705A inhibits the insulin-like growth factor-1 receptor and anaplastic lymphoma kinase and shows antitumour activity in experimental models of human cancers. *Mol Cancer Ther* 2009;8:2811-20.

36. Sabbatini P, Rowand JL, Groy A, Korenchuk S, Liu Q, Atkins C, et al. Antitumour activity of GSK1904529A, a small molecule inhibitor of the insulin-like growth factor-I receptor kinase. *Clin Cancer Res* 2009;15:3058-67.

37. Bertrand FE, Steelman LS, Chappell WH, Abrams SL, Shelton JG, White ER, et al. Synergy between an IGF-1R antibody and Raf/MEK/ERK and PI3K/Akt/mTOR pathway inhibitors in suppressing IGF-1R-mediated growth in hematopoietic cells. *Leukemia* 2006;20:1254-60.

38. Baumann P, Hegemeier H, Mandl-Weber S, Franke D, Schmidmaier R. Myeloma cell growth inhibition is augmented by synchronous inhibition of the insulin-like growth factor-1 receptor by NVP-AEW541 and inhibition of mammalian target of rapamycin by Rad001. *Anticancer Drugs* 2009;20:259-66.

39. Karp DD, Paz-Ares LG, Novello S, Haluska P, Garland L, Cardenal F, et al. Phase II study of the anti-insulin-like growth factor type 1 receptor antibody CP-751,871 in combination with paclitaxel and carboplatin in previously untreated, locally advanced, or metastatic non-small-cell lung cancer. *J Clin Oncol* 2009;27:2516-22.

40. Tolcher AW, Sarantopoulos J, Patnaik A, papadopoulos K, Lin C, Rodon J, et al. Phase I, pharmacokinetic, and pharmacodynamic study of AMG 479, a fully human monoclonal antibody to insulin-like growth factor receptor 1. *J Clin Oncol* 2009;27:5800-7.

41. Hidalgo M, Tirado Gomez M, Lewis N, Vuky JL, Taylor G, Hayburn JL, et al. A phase I study of MK-0646, a humanized monoclonal antibody against insulin-like growth factor receptor type 1 (IGF1R) in advanced solid tumour patients in a q2wk schedule. *J Clin Oncol* 2008;26 Suppl 15:3520.

42. Higano Cs, Yu SH, Gordon MS, LoRusso P, Fox F, Katz TL, et al. A phase I, first in man study of weekly IMC-A12, a fully human insulin like growth factor-1 receptor IgG1 monoclonal antibody, in patients with advanced solid tumours. *J Clin Oncol* 2007;25 Suppl 18:3505.
43. Olmos D, Postel-Vinay S, Molife LR, Okuno SH, Schuetze SM, Paccagnella ML, et al. Safety, pharmacokinetics, and preliminary activity of the anti-IGF-1R antibody figitumumab (CP-751,871) in patients with sarcoma and Ewing's sarcoma: a phase 1 expansion cohort study. *Lancet Oncol* 2010;11:129-35.
44. Kurzrock R, Patnaik A, Aisner J, Warren T, Leong S, Benjamin R, Eckhardt SG, et al. A phase I study of weekly R1507, a human monoclonal antibody insulin-like growth factor-I receptor antagonist, in patients with advanced solid tumours. *Clin Cancer Res* 2010;16:2458-65.
45. Attard G, Fong PC, Molife R, Reade S, Shaw H, Reid A, et al. Phase I trial involving the pharmacodynamic (PD) study of circulating tumour cells, of CP-751,871 [®], a monoclonal antibody against the insulin-like growth factor 1 receptor (IGF-1R) with docetaxel (D) in patients (p) with advanced cancer. *J Clin Oncol* 2006;24 Suppl 18:3023.
46. Lacy MQ, Alsina M, Fonseca R, Paccagnella ML, Melvin CL, Yin D, et al. Phase I, pharmacokinetic and pharmacodynamic study of the anti-insulinlike growth factor type 1 Receptor monoclonal antibody CP-751,871 in patients with multiple myeloma. *J Clin Oncol* 2008;26:3196-203.
47. Atzori F, Tabernero J, Cervantes A, Botero M, Hsu K, Brown H et al. A phase I, pharmacokinetic (PK) and pharmacodynamic (PD) study of weekly (qW) MK-0646, an insulin-like growth factor-1 receptor (IGF1R) monoclonal antibody (MAb) in patients (pts) with advanced solid tumours. *J Clin Oncol* 2008;26:Suppl 15:3519.
48. Hidalgo M, Tirado Gomez M, Lewis N, Vuky JL, Taylor G, Hayburn JL, et al. A phase I study of MK-0646, a humanized monoclonal antibody against the insulin-like growth factor receptor type 1 (IGF1R) in advanced solid tumour patients in a q2 wk schedule. *J Clin Oncol* 2008;26 Suppl 15:3520.
49. Lindsay CR, Chan E, Evans TR, Campbell S, Bell P, Stephens AW, et al. Phase I dose escalation study of continuous oral dosing of OSI-906, an insulin like growth factor-1 receptor (IGF-1R) tyrosine kinase inhibitor, in patients with advanced solid tumours. *J Clin Oncol* 2009;27 Suppl 15:2559.
50. Patel S, Pappo A, Crowley J, Reinke D, Eid J, Ritland S, et al. A SARC global collaborative phase II trial of R1507, a recombinant human monoclonal antibody to the insulin-like growth factor-1receptor (IGF1R) in patients with recurrent or refractory sarcomas *J Clin Oncol* 2009;27 Suppl 18:10503.
51. Pappo AS, Patel S, Crowley J, Reinke DK, Staddon AP, Kuenkele K, et al. Activity of R1507, a monoclonal antibody to the insulin-like growth factor-1 receptor (IGF1R), in patients (pts) with recurrent and refractory Ewing's sarcoma family of tumours (ESFT): results of a phase II SARC study. *J Clin Oncol* 2010;28 Suppl 15:10000.
52. Tap WD, Demetri GD, Barnette P, Desai J, Kavan P, Tozer R, et al. AMG 479 in relapsed or refractory Ewing's family tumours (EFT) or desmoplastic small round cell tumours (DSRCT): Phase II results. *J Clin Oncol* 2010;28 Suppl 15:10001.
53. Reidy DL, Vakiani E, Fakih MG, Saif MW, Hecht JR, Goodman-Davis N, et al. Randomized, phase II study of the insulin-like growth factor-1 receptor inhibitor IMC-A12, with or without cetuximab, in patients with cetuximab- or panitumumab-refractory metastatic colorectal cancer. *J Clin Oncol* 2010;28:4240-6.
54. Higano C, Alumkal J, Ryan CJ, Yu EY, Beer Tm, Chandrawansa K, et al. A phase II study evaluating the efficacy and safety of single agent IMC A12, a monoclonal antibody(MAb),againsttheinsulin-likegrowthfactor-1receptor (IGF-IR), as monotherapy in patients with metastatic, asymptomatic castration-resistant prostate cancer (CRPC). *J Clin Oncol* 2009; 27 Suppl 15:5142.
55. Chi KN, Gleave ME, Fazli L, Goldenberg SL, So A, Kollmannsberger CK, et al. A phase II of preoperative figitumumab (F) in patients (pts) with localized prostate cancer (Pca). *J Clin Oncol* 2010;28 Suppl 15:4662.
56. Shah S, Small E. Emerging biological observations in prostate cancer. *Expert Rev Anticancer Ther* 2010;10:89-101.
57. Karp DD, Paz-Ares LG, Novello S, Haluska P, Garland L, Cardenal F, et al. Phase II study of the anti-insulin-like growth factor type 1 receptor antibody CP-751,871 in combination with paclitaxel and carboplatin in previously untreated, locally advanced or metastático non-small-cell lung cancer. *J Clin Oncol* 2009;27:2516-22.
58. Kindler HL, Richards DA, Stephenson J, Garbo LE, Rocha Lima CS, Safran H, et al. A placebo-controlled, randomized phase II study of conatumumab (C) or AMG 479 (A) or placebo (P) plus gemcitabine (G) in patients (pts) with metastatic pancreatic cancer (mPC). *J Clin Oncol* 2010;28 Suppl 15:4035.
59. Jassem J, Langer CJ, Karp DD, Mok T, Benner RJ, Green SJ, et al. Randomized, open label, phase III trial of figitumumab in combination with paclitaxel and carboplatin versus paclitaxel and carboplatin in patients with non-small cell lung cancer (NSCLC) *J Clin Oncol* 2010;28 Suppl 15:7500.
60. Kaufman PA, Ferrero JM, Bourgeois H, Kennecke H, De Boer R, Jacot W, et al. A Randomized, Double-Blind, Placebo-Controlled, Phase 2 Study of AMG 479 With Exemestane (E) or Fulvestrant (F) in Postmenopausal Women With Hormone-Receptor Positive (HR+) Metastatic (M) or Locally Advanced (LA) Breast Cancer (BC). S1-4. *SABC* 2010.